



Management of severe acute pancreatitis in 2019

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Abstract: Pancreatic fluid collections (PFCs) are frequent complications in severe acute pancreatitis that are the result of damage to the pancreas to include but not limited to trauma, surgery, autoimmune diseases, alcohol abuse, infections, medications, gallstones, metabolic disorders, and premalignant or malignant conditions. The majority of these collections resolve spontaneously; however, if the collection is infected or causes symptoms to include abdominal pain, nausea, vomiting, diarrhea, fevers, and tachycardia, drainage is indicated. Drainage of PFCs can be accomplished surgically, percutaneously, or endoscopically and should be approached in a multidisciplinary fashion for best overall patient care and outcomes. Before the introduction of endoscopic procedures, surgical and percutaneous drainage was the preferred modality. Today a minimally-invasive “step-up” approach is generally accepted depending upon the specific characteristics of the PFC and clinical presentation. Endoscopic ultrasound-guided PFC drainage is favored due to high success rates, shorter hospital stays, and lower cost. Direct debridement of walled-off pancreatitis can now be performed endoscopically with higher success rates with larger caliber fully covered metal stents. At large, the field of endoscopic techniques has evolved, and more specifically, the management of PFCs continues to evolve with increasing experience and with the advent of new stents and accessories, leading to increased efficacy with less adverse events.

Keywords: Pancreatitis; pancreatic fluid collections (PFCs); pancreatic necrosis; endoscopic drainage of pancreatic fluid collections; endoscopic ultrasound guided drainage of pancreatic fluid collections

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Introduction

Acute pancreatitis is described as two or more of the following criteria: abdominal pain suggestive of pancreatitis, serum amylase or lipase level greater than three times the upper limit of normal and characteristic imaging findings (1). Mild acute pancreatitis, or interstitial pancreatitis, is characterized by absence of organ failure or systemic complications. Moderately severe acute pancreatitis is characterized by transient organ failure (resolving within 48 hours) and/or local systemic complications without persistent organ failure (>48 hours). Severe acute pancreatitis is characterized by persistent organ failure that may involve one or more organ systems. This includes

shock (systolic blood pressure <90 mmHg), pulmonary insufficiency ($\text{PaO}_2 \leq 60$ mmHg), acute kidney injury (serum creatinine level >2 mg/dL), and gastrointestinal bleeding. Approximately 15–25% of all patients with acute pancreatitis develop severe pancreatitis with local complications, such as necrosis, pseudocyst, or abscess formation (2).

Nutrition

Patients who develop pancreatitis are often placed NPO until the inflammation has resolved and can withstand an oral diet. Current literature supports the notion that enteral nutrition is superior to parenteral nutrition. In severe acute

pancreatitis a feeding strategy is advised for the patient to eat food orally and when patient cannot tolerate an oral diet after 72 hours, enteral feed are often started (3).

Scoring systems

The ability to predict the severity of pancreatitis may help identify patients at an increased risk for morbidity and mortality. Clinical predictors of severe acute pancreatitis include age >55, alcoholic pancreatitis, the interval to onset, obesity (body mass index >30), and organ failure (4). There are several scoring systems, although most are imperfect due to the tedious nature required for calculation. Glasgow and Ranson scores take 48 hours to complete, can be used only once, and have a low sensitivity and specificity (5). The APACHE II score (Acute Physiology and Chronic Health Examination) has 12 physiologic measures and extra points based upon age and presence of chronic disease. The BISAP (Bedside Index for Severity in Acute Pancreatitis) score can be utilized to identify patients of high risk for 120-day mortality or severe disease in acute pancreatitis based upon each present criterion: BUN >25 mg/dL, acute change in mental status, systemic inflammatory response syndrome (SIRS), age >60 years, and pleural effusions (6). Compared with the Ranson criteria and APACHE II score, the BISAP score outperformed in specificity (91%), but had suboptimal sensitivity (56%) for mortality and severe acute pancreatitis within 3 months (7). Radiological imaging can also be used to determine the severity of acute pancreatitis utilizing the CT severity index (CTSI). The CTSI sums the Balthazar score grading of pancreatitis (extent of pancreatic inflammation and the presence of fluid collections) and CT scan grading of pancreatic necrosis with a sensitivity of 92% and specificity of 100% (8,9).

Classification of pancreatic fluid collections

Conclusive research in pathophysiologic understanding and improvement in diagnostic tools initiated a revision of the Atlanta Criteria in 2012 (10). The revised classification divided acute pancreatitis (4 weeks after pancreatitis episode) into two distinct subtypes, interstitial edematous pancreatitis (IEP) and necrotizing pancreatitis. Interstitial edematous acute pancreatitis is characterized by acute inflammation of the pancreatic parenchyma and peripancreatic tissues, but without tissue necrosis. IEP can lead to acute pancreatic fluid collections (<4 weeks) or pseudocysts (>4 weeks) (11). Necrotizing acute pancreatitis is characterized by inflammation

associated with pancreatic parenchymal necrosis and/or peripancreatic necrosis. Necrotizing pancreatitis can lead to acute necrotic collections (<4 weeks) and walled off pancreatic necrosis (WOPN, <4 weeks) (11). The new subdivisions of acute pancreatitis are essential since the management and treatment are tailored to the type of collection present (11).

Procedural considerations for drainage

Up to 80% of PFCs resolve spontaneously and at least 25% will decrease in size with time (12). When the PFCs increase in size, they often become symptomatic and require drainage (13). Indications for drainage include infection, abdominal pain, vomiting, early satiety, persistent jaundice, and weight loss. If drainage is required and the patient is stable, it is suggested that it be performed >4 weeks to allow for encapsulation (14).

The modality of the drainage is dependent on the type of fluid collection, and the method for drainage has evolved with time. Effective management often requires collaboration with a multidisciplinary team of surgeons, radiologists, and gastroenterologists. Before the advancement of endoscopic technology and techniques, surgery was the preferred method. Surgical drainage of PFCs (cystogastrostomy) is performed by creating a tract between the PFC and the stomach or small bowel and has excellent resolution rates (91–97%) (15). However, when external drainage of pancreatic necrosis and necrosectomy is required, there is a high failure rate of 20% to 30% and a complication rate of 4% to 30% including pancreatic duct leaks and abscesses (16).

Percutaneous drainage is an effective intervention in fragile patients, with severe comorbidities or those with immature infected fluid collections who are not candidates for surgical or endoscopic interventions. The limitations of percutaneous drainage include the lack of a safe access route, hemorrhage, and pancreatic fistulas (17). The patients also often require more interventions and have longer hospitalizations (18).

PFCs typically occupy free space between anatomic structures including the stomach and duodenum. The close proximity has promoted the development of endoscopic techniques for drainage with fewer complications (19). For consideration of endoscopic drainage, a PFC must measure at least 3 cm in size, INR <1.5, platelets >50,000/ μ L, and discontinuation of direct oral anticoagulants for at least 48 hours (20). In the past, conventional endoscopic drainage

was performed when a bulge was seen in the lumen of the GI tract. With the advent of endoscopic ultrasound, the fluid collection can be fully characterized in terms of size, wall thickness and regional vasculature.

Several studies demonstrated that a minimally invasive “step up” approach decreased mortality and complications as compared to open surgical interventions (21,22). The shift to endoscopic therapy as the preferred method of PFC management was propelled by a randomized comparative trial by Varadarajulu *et al.* comparing surgical *vs.* endoscopic approach to PFC treatment. It was determined that though the two approaches had similar rates of PFC resolution and complications, the total mean cost was lower for patients managed by endoscopy than surgery (23). Since then, various studies evaluated the comparison and overall outcomes between surgical (SD), percutaneous (PD) and endoscopic drainage (ED). Morton *et al.* compared SD and PD and concluded that SD has fewer complications, less inpatient mortality, reduced hospital stay, and overall cost (24). Larg and colleagues compared surgical versus endoscopic cystgastrostomy and necrosectomy. The success rate was 93.3% and 90% in the laparoscopic and endoscopic groups, respectively (25). Overall, the cost, length of hospital stay, and invasiveness is higher with the surgical approach when compared to an endoscopic approach (26). A study by Bakker *et al.* demonstrates endoscopic necrosectomy reduces the proinflammatory response compared with surgical necrosectomy (27). However, van Brunschot recently suggested that the step up approach in infected necrotizing pancreatitis by endoscopy was not superior to a surgical approach in reducing major complications (bleeding, perforation) or death (28). Therefore, ongoing evolution of the endoscopic technique requires attention to be focused on decreasing the risk of bleeding associated with regional vasculature and successful stent deployment. Keane and colleagues completed a study between the comparison of ED *vs.* PD for the treatment of pancreatic fluid collections. The study demonstrated a low risk of stent migration, pneumoperitoneum, esophageal perforations, pneumothorax and aspiration pneumonia with endoscopic drainage (29). However, percutaneous drainage was considered inferior due to residual collections (20% *vs.* 53%) and the need for reintervention or surgical drainage (4% *vs.* 6%) (26). While there is not a consensus on how to treat these patients as a whole, each case should be approached in an as minimally invasive manner as possible, and the step up approach is generally accepted.

Methods

History of endoscopic interventions of pancreatic fluid collections

The first endoscopic aspiration was in 1975 with 21 gauge needle drainage in a 31-year-old patient with a pancreatic fluid collection secondary from alcoholic pancreatitis (30).

Ten years later, PFC drainage was reported by the creation of a fistulotomy (31). The hypothesis was that rather than simply aspirating the collection that yielded insufficient drainage, the creation of a larger opening would lead to resolution. Fistulotomies were limited by the rapid, spontaneous closure of the opening before complete cyst resolution, resulting in recurrence and superinfection (32). A nasocystic drainage catheter with normal saline irrigation was utilized to provide sustained drainage, but this technique is cumbersome with poor tolerance of the nasocystic catheter and inferior anchoring methods.

Since superior endoscopic methods were in demand, internal stent placement was performed an alternative to nasocystic catheter drainage (33,34). The stent maintains patency of the fistula without the need for external drainage and patient discomfort. In 1993, 7 or 10 Fr plastic biliary stents for transmural PFC drainage were first reported, with success rates exceeding 90% (33,34).

Prior to EUS, conventional drainage was performed by evaluating the luminal wall for a bulge (35). Diagnostic puncture was made into the bulge with aspiration followed contrast injection to localize the PFC. and then immediate wire placement as a one-step procedure. A guidewire is positioned within the collection with subsequent dilation and stent placement.

Technique

The endoscopist should have a strong understanding of both endoscopic ultrasound and ERCP techniques. Patients undergo general anesthesia with intubation for airway management and prevention of aspiration, and all patients receive antibiotics during the procedure. Using a linear array echoendoscope, the fluid collection is localized from the stomach or small intestine. Color flow doppler is used to identify regional vasculature prior to puncture. The distance between the gastrointestinal lumen and the fluid collection should be measured to ensure an optimal puncture site prior to drainage. A 19-gauge FNA needle is used to puncture the stomach or duodenal wall under direct visualization

by EUS. The stylet is removed and fluid can be aspirated to send for culture. A guidewire is then advanced through the needled and is coiled within the fluid collection under endosonographic and/or fluoroscopic guidance. The needle is exchanged for a dilating catheter or balloon dilator to create a fistula and facilitate stent placement. This step can be challenging due to positioning and chronic inflammatory changes of the GI tract wall. If necessary, an electrocautery device such as a needle knife or cystostome can be used to create the fistula followed by adequate dilation to 6–8 mm. Depending on the type of fluid collection, an appropriate stent is then deployed.

Stent selection

If the fluid collection is a pure pseudocyst, plastic double pigtail stents (PS) are placed as standard practice due to accessibility and low cost (36). Multiple stents can be placed across the fistula to enhance drainage and minimize complications from stent occlusion. This can result in an increase in procedure time, so the use of self-expandable metal stents (SEMS) was described. SEMS create an alternative to balloon catheter dilation of the cystenterostomy tract, and is preferred if there is necrotic material within the collection. The larger lumen of the SEMS improves drainage, and reduces rates of stent occlusion, recurrence, and secondary infections (37). SEMS reduce the risk of leakage between the cyst and enteric lumens, reduce the risk of bleeding by creating a tamponade effect, and have shorter procedure times (37). At the discretion of the endoscopist, one or more double pigtail plastic stents can be placed within the metal stent, which serves as an anchor to prevent stent migration and perforation of the collection wall as it is resolving (38). When comparing the use of plastic stents versus FCSEMS in pancreatic fluid collections, there were no significant difference in success (85% *vs.* 83%) (39). Overall outcomes and cost effectiveness of SEMs are superior in comparison to plastic stents as seen in a study by Sharaiha *et al.* where complete resolution of PFCs using double pigtail plastic stent was lower compared to drainage with FCSEMSs (89% *vs.* 98%; $P=0.01$) (40). Procedural adverse events were noted in 31% in the plastic stent group and 16% in the FCSEMS group ($P=0.006$) (40,41).

When endoscopic debridement of a necrotic collection is necessary or if multiple sessions are required for debridement, the stent must be removed first in order to advance the scope into the necrotic cavity. To overcome

this limitation, fully covered esophageal self-expanding metal stents (FCESEMS) have been described to provide superior drainage. The large caliber diameter allows for a gastroscope to be advanced into the collection for direct endoscopic irrigation and debridement with various accessories. Sarkaria *et al.* described this technique for 17 patients with an 88% success rate and adverse event rate of 5% (perforation of fistula tract during dilation, which was managed conservatively) (42).

Tubular stents, whether plastic or metal, have several design flaws for transluminal drainage. There is no mechanism to anchor the collection to the lumen of the GI tract. Thus, there is concern for leakage of contents and premature stent migration, which can require surgical intervention or can be fatal (43). Tubular stents are often longer than what is required. The exposed end of the stent may cause tissue trauma resulting in bleeding or perforation (43). The length of tubular stents also causes a predisposition to clog from cyst debris or food residue.

As endoscopic drainage for pancreatic fluid collections has evolved, experienced endoscopists are aware that dedicated stents are needed facilitate drainage. A lumen apposing stent (LAMS) for transluminal drainage was designed to address the specific concerns with tubular metal stents. Several lumen-apposing metal stents (LAMS), have been designed including the AXIOS stent (Boston Scientific, Marlborough, MA United States), Nagi stent (Taewoong Medical Co, Ilsan, South Korea), and Niti-S SPAXUS stent (TaeWoong Medical Co., Ltd., Ilsan, South Korea) (44).

The AXIOS stent was designed in 2004 and is the most commonly utilized LAMS in the US. The AXIOS is a nitinol braided FCSEMS with bilateral double-walled flanges in a dumbbell configuration that are vertical to the lumen and appose tissue walls to create an anastomosis (37). The respective flanges are designed to reduce stent migration and approximate structures to reduce rates of perforation, leak, and stent erosion (45). These stents have varying diameters, including 10, 15, and 20 mm that promote superior drainage and allow for direct endoscopic necrosectomy (46).

The stents are deployed in a similar fashion as other SEMS. Once the fistula tract has been dilated, the stent is advanced over the guidewire and into the fluid collection. The distal flange of the LAMS is then deployed under endosonographic guidance. The deployment catheter is then gently retracted so that the flange is secured against the wall of the fluid collection. The proximal flange can be deployed in the echoendoscope working channel, and is then released by

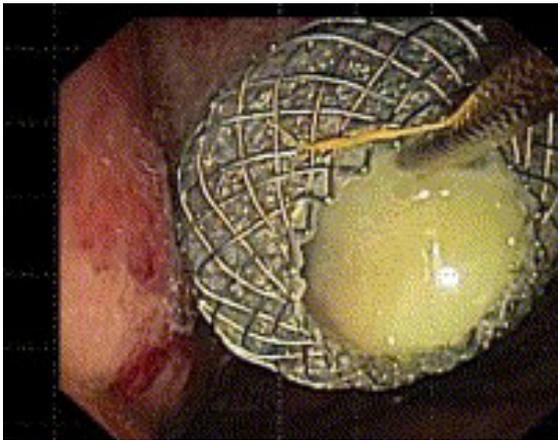


Figure 1 Axios stent draining pancreatic fluid collection with purulent drainage.

advancing the catheter hub while simultaneously retracting the echoendoscope away from the bowel wall (*Figure 1*).

The catheter sheath also has two radiopaque markers indicating each end of the preloaded stent to enable fluoroscopic control of stent position, but the use of fluoroscopy for routine stent deployment is not needed.

Due to the complex nature of each step of EUS guided pancreatic fluid drainage and stent deployment, endoscopists are in need of accessories to streamline this process and reduce the risk for technical complications. In 2013, “hot” AXIOS, the Axios stent with an electrocautery-enhanced delivery system was developed. This system enables the operator to directly access the lumen of the collection with a LAMS-loaded catheter followed by immediate deployment of the stent. The electrocautery-enhanced delivery system reduces the risk of leakage since the tract remains sealed off after puncture by the 10.8 F delivery catheter until the stent is deployed, eliminating exchanging various catheters over a wire (47). The tamponade effect of the fully covered LAMS reduces the risk of bleeding from the transmural tract (48). LAMS is superior to plastic stenting in the setting of WON requiring endoscopic drainage with increased clinical success (LAMS, 80.4% and PS 57.5% ($P=0.001$), shorter procedure time (LAMS 50.4 ± 26.5 vs. PS 64.6 ± 34.0 minutes, $P=0.003$), decreased need for surgery (16.1% PS vs. 5.6% LAMS, $P=0.02$), and lower rate of recurrence with a comparable rate of adverse events (46).

Endoscopic necrosectomy

Endoscopic necrosectomy is the endoscopic manual

debridement of tissue from walled off necrotic pancreatic collections. The scope can be advanced directly into the collection and water irrigation and suction are utilized to clear the cavity. Larger pieces of necrotic material can be removed with various accessories including forceps, standard polypectomy snare, Roth nets or a trapezoid basket (49). Lavage with 3% diluted hydrogen peroxide can help to facilitate debridement (50). Discontinuing proton pump inhibitors may foster auto-digestion of the necrotic tissue by gastric acid (51). Nasocystic drains can be used as an adjunct to treatment with the intermittent irrigation of the WON with normal saline to decrease the interval or decrease the need for repeat endoscopic debridement. However, the clinical success of endoscopic transmural drainage using the adjunct of nasocystic drain for PFCs may depend on the presence of necrotic material within the cavity (52). Necrosectomy sessions can be performed every 2–5 days until the majority of non-adherent necrotic material is removed, and/or there is clinical improvement (53). The timing for removal is evaluated between 1–2 months after the initially placement. During this time period the repeat cross-sectional imaging is performed to evaluate for resolution of the pancreatic fluid collection. The stent can then be removed if there is complete resolution of the fluid collection (53).

Variations of this technique are performed based on the clinical status of the patient and the morphologic appearance of the collection. Plastic stents can be utilized, but each session will require that several stents be removed with fistula tract dilation to facilitate advancement of the scope into the collection. After the debridement session is completed, additional stents are placed to keep the fistula patent. Due to the tedious nature of the procedure with the sole use of plastic double pigtail stents, it has been more common placed to use fully covered metal stents as discussed above, most commonly the lumen apposing metal stents. Studies have demonstrated excellent efficacy and safety, so LAMS are quickly becoming favored over plastic stents due to better clinical success rates and few adverse events.

Several techniques have been described in order to increase the efficacy of endoscopic drainage of PFCs (54). In difficult cases requiring multiple endoscopies for repeat necrosectomy a percutaneous drain may be placed to help maintain adequate drainage (55). Varadarajulu *et al.* described the use of “Gateway” drainage to decrease the need for surgery and endoscopic necrosectomy by utilizing 2 or 3 transmural tracts created by using EUS guidance

between the necrotic cavity and the GI lumen. One tract is used to flush normal saline solution through a nasocystic catheter and multiple stents are used to increase drainage of necrotic contents. Plastic stents can be used in the setting of large collections to obtain access to deeper portions of the pancreatic fluid collection to facilitate resolution (49). Navarrette *et al.* described the use of a percutaneous drain as an access point for difficult to access collections that are close to the skin. A fully covered esophageal metal stent is placed through the skin and across the percutaneous fistula tract into the collection for endoscopic necrosectomy (55).

Role of ERCP

Approximately 37–67% of patients with acute pancreatitis will have a pancreatic duct injury (56). Pancreatic duct disruptions can result from acute or chronic pancreatitis, trauma, or surgical injury and the role of ERCP can be successful in detecting this presence with the help of a multidisciplinary approach when considering stenting the PD duct to stop or prevent recurrent collections (57). Trevino and colleagues have shown that transpapillary stenting can lead to successful resolution of pancreatic duct leaks, particularly in partial duct disruptions (58). Shrode *et al.* demonstrated that there is limited benefit for endoscopic stenting in complete ductal disruptions, especially if the disruption cannot be bridged with the stent (59).

Adverse events

The reported rate of adverse events associated with EUS guided drainage of fluid collections ranges from 4–21%, including bleeding, infection, perforation and stent migration (60). The introduction of LAMS for PFC drainage has substantially decreased the risk of adverse events, but there are reports of bleeding, stent maldeployment/migration, and buried stents due to a long course (61). Careful patient selection, with appropriate assessment of the size, components and location of the collection, and an ability to predict complications are needed to improve the safety of these stents.

Role in clinical practice

The drainage of PFCs should be reserved for advanced gastroenterologists who have expertise in interventional EUS and ERCP. ASGE guidelines recommend that endoscopic drainage of PFCs be performed only when collections are symptomatic, infected, stagnate collections >8 weeks, or rapidly enlarging cysts after maturation of the

cyst wall of PFCs bulge (62). ESGE guidelines recommend performing endoscopic or percutaneous drainage of infected walled-off necrosis as the first interventional method. If there is no improvement following endoscopic transmural drainage of walled-off necrosis, endoscopic necrosectomy or minimally invasive surgery (percutaneous drainage) is preferred over open surgery as the next therapeutic step.

Future considerations

The endoscopic treatment of severe pancreatitis with pancreatic fluid collections has evolved dramatically over the past 10 years. With variations of techniques and the advancement of accessories, the overall success rates will climb. Ongoing research and development of dedicated accessories and tools for drainage and debridement are necessary in order to improve efficacy and decrease adverse event rates.

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