Introduction

Pancreatic ductal adenocarcinoma (PDAC) is the most common pancreatic neoplasm with 5-year survival as low as 6% (1,2). The low 5-year survival rate is often due to late diagnosis and only 20% of the patients are eligible for surgical resection (2). Over the past two decades, advances have been made in molecular genetics, pharmaco-oncology and peri-operative care. However, survival rates remain dismal due to the lack of screening modalities, advanced stage of disease at presentation and aggressive tumour biology (3). It is therefore imperative to explore potential treatment avenues to improve survival in these groups of patients. Anti-estrogenic hormone therapy (AEHT) is a potential treatment alternative which has demonstrated improved recurrence-free survival in breast cancer in estrogen receptor (ER) positive subgroups (4), and is well-tolerated by patients (5).

Sex hormone receptors are expressed in a myriad of
cancers, including pancreatic cancer (6). ER is a type of sex hormone receptor which have been reported to be expressed in pancreatic cancer, though there have been inconsistencies in their detection (7,8). Presence of ER expression in PDAC may potentially guide treatment with AEHT. Several studies have been conducted to explore the role of AEHT (mainly tamoxifen) in the management of PDAC; however, results are equivocal with varying treatment response across different studies (9-16). It has been postulated that the inconsistent results may be attributed to the lack of assessment of estrogen dependence in PDAC (17). Hence, this study aims to identify the presence of ER in PDAC specimens to guide potential use of AEHT in the management of unresectable PDAC.

Methods

This is a retrospective case control study of 10 patients (5 males, 5 females) who underwent pancreatic resections for PDAC. A list of patients diagnosed with PDAC as per the ICD-10 (International Classification of Diseases, 10th Revision) codes and who underwent pancreatic resections from 2011 to 2012 were generated from the lead authors operative log (18). Five male and 5 females were randomly selected from the list of patients generated. It has been well-described in literature that a minimum sample size of 10–12 for pilot studies provides the ability to test a hypothesis (19-21). Presence of ER expression in ≥2 of the 10 PDAC specimens will clinically justify further studies involving the use of AEHT in the management of PDAC. Post-operative specimens were prepared for a minimum of seven years by the Department of Pathology in our institution. Specimens with inadequate tissue volume were excluded from the study. Upon completion of the patient selection, sections of the post-operative specimens were prepared and sent for ER staining. This study was approved by the hospital institutional review board (reference number: 2012/00919) and funded by the National Research Foundation Scholarship Grant.

Specimen preparation

Sections of pancreatic cancer including adjacent normal pancreatic tissue were cut from formalin fixed, paraffin embedded tissue blocks and stained for ER using Novocastra Liquid Mouse Monoclonal Antibody Estrogen Receptor™ (Product code: NCL-L-ER-6F11) (Leica Biosystems, Newcastle Upon Tyne, UK) and Ventana OptiView DAB IHC Detection Kit™ (Ventana Medical Systems, Arizona, USA) with appropriate tissue controls. Pancreatic tissue specimens that were analysed included (I) ductal epithelial cells; (II) acinar cells; (III) islet cells; (IV) intralobular stromal cells; and (V) adenocarcinoma cells. Reporting of ER staining was performed in accordance to ER staining for breast cancer specimens (22): (I) proportion of cells (0–100%) with ER positive nuclear staining and (II) intensity of staining (weak, moderate and strong). Specimen preparation and histopathological analysis were performed by a single, blinded pathologist.

Results

Intralobular stromal cells were positively stained for ER in 7/10 (70%) of the cases; these were stromal spindly cells, all of which had nuclear ER-positive staining were of weak intensity and were patchy in distribution (Figure 1). Islet cells (<1%) stained for ER in 3/10 (30%) of the cases (Figure 2). Ductal epithelial cells, acinar cells and adenocarcinoma cells stained negative for ER in all of the cases (Figure 3). ER positive cells were noted in both normal pancreatic tissue and areas of chronic inflammation adjacent to the pancreatic adenocarcinoma cells.

Discussion

In this pilot study, PDAC did not express ER. ER is present in some of the normal pancreatic tissues (intralobular stromal cells 7/10, islet cells 3/10). To our knowledge, current studies do not show expression of ER limited to intralobular stromal cells and islet cells. It is possible that islet cells may have been incorrectly identified due...
to lack of confirmatory staining with anti-insulin or anti-glucagon antibodies. However, it has described in literature that the use of antibody-staining techniques may not be necessary (23); islet cells can be easily identified with background counter stain by a trained pathologist. Negative staining for ER in ductal epithelial and acinar cells may be due to the true absence of ER or very low quantities of ER which is undetectable by the immunohistochemical technique.

It is still uncertain whether the ER expression is present in all patients with PDAC. A study by Greenway et al. failed to demonstrate presence of ER receptors at five different sites in a patient with underlying PDAC (7). Our pilot study is also supported by the studies by Singh et al. (8), and Taylor et al. (24), which detected little to no ER expression by immunohistochemistry. The lack of ER staining in our pilot study is unlikely due to the specimen preparation and immunohistochemistry analysis: adequate tissue controls were used to identify problems with tissue fixation with formalin; this is of important significance as it is widely documented that ER is labile and good fixation is required to identify its presence (25). Use of immunohistochemistry for ER performed formalin fixed paraffin embedded tissue has also been documented to be a reliable and convenient method to study ER expression (26).

The role of AEHT in PDAC is still uncertain. Existing studies thus far show equivocal evidence on its use in the management of unresectable PDAC. A study by Wong et al. demonstrated that tamoxifen prolongs survival in postmenopausal women \((n=6/15, 3\) with minimum 2-year survival) with unresectable PDAC (12). However, other randomized studies fail to demonstrate the survival benefit of tamoxifen in unresectable pancreatic adenocarcinoma (15,24). The lack of survival benefit in patients treated with tamoxifen could be attributed to the lack of ER expression, as demonstrated in the study by Taylor et al. (ER expression positive in \(n=0/25\)) (24). It was traditionally postulated that the role of tamoxifen in breast cancer is due to its effect on nuclear ERs (27). However, a recent translational study by Cortes et al. in 2019 on mice demonstrated that a new mechanism of action of tamoxifen which is independent of its effect on nuclear ERs (28); this involves a mechanical downregulation of hypoxia-inducible factor-1 alpha (HIF-1A) which reduces the adaptive response of PDAC to hypoxia and may be responsible for reprogramming of the tumour microenvironment and its therapeutic role in some of studies which documented survival benefit of tamoxifen in PDAC. Other mechanisms of action on the role of tamoxifen in cancer treatment have also been described (29).

The presence of ER expression in other organs have also guided several investigations on the role of anti-hormonal therapy (6). However, these results have been dismal. A randomized control trial on the use of tamoxifen in hepatocellular carcinoma (HCC) by the Cancer of the Liver Italian Programme Group in 1998 (30), and a multicentre regional randomized control trial by Chow et al. in 2002 demonstrated the lack of survival benefits in HCC (31); the latter involved the use of high-dose tamoxifen which inhibits HCC via ER-independent mechanisms.

One of the limitations of this study is that only a single pathologist was involved in the histopathological analysis of the specimens. However, it has been demonstrated that a second opinion surgical pathology is unlikely to change diagnosis with an overall incidence of 1.4% of

Figure 2 Islet cells showing occasional nuclear estrogen receptor (ER) positivity (black arrow), \(\times400\) magnification.

Figure 3 Pancreatic ductal adenocarcinoma cells showing lack of estrogen receptor (ER) staining, \(\times100\) magnification.
changed diagnoses, and an even lower incidence of changed diagnosis for gastrointestinal (1.2%) and endocrine (0%) pathologies (32). Due to the small sample size in this pilot study, this is likely to be unremarkable. In addition, this is a pilot study with a small sample. A larger sample would be ideal for detecting the presence of ER staining but this is limited by inadequate funding to permit ER staining for all the patients who underwent pancreatic resections for PDAC. Furthermore, a sample size of ten was determined to be acceptable for a pilot study (19-21).

In conclusion, this pilot study did not detect the presence of ER expression in PDAC. Hence, we were unable to proceed to explore AEHT role in management of ER-positive PDAC. The role of AEHT in pancreatic cancer remains uncertain and does not appear to be of value at present. Our study demonstrates the presence of ER expression in intralobular stromal and islet cells, which was previously unreported. Further studies are required to determine the significance of weak and patchy ER expression in intralobular stromal and islet cells.

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Footnote

Conflicts of Interest: The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. This study was approved by the hospital institutional review board (reference number: 2012/00919).

References


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